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PREPARATION OF METAL OXIDE SUPPORTS LOADED WITH BIOMOLECULES

The invention relates to a method for the preparation of metal oxide supports loaded with biomolecules, to the thus obtained metal oxide supports, to the use of said supports for performing probe-based assays and to a kit of parts comprising said support and a detection means.

The present invention specifically relates to a method for the preparation of metal oxide supports loaded with biomolecules, comprising the steps of: (a) activating the surface of the support by means of an silanating agent comprising an amine group; and (b) loading the support by attaching biomolecules to the activated surface.

Such a method is known, e.g. from WO 99/002266, in which method aluminium oxide membranes are activated using 3-aminopropyl triethoxysilane (APS) after which oligonucleotide probes are covalently coupled to the activated membranes. A similar method is described for aluminiumoxide nanoparticles carrying antigens (Bioconjugate Chem. 1997, 8, 424-433). Further, for chromatographic purposes, aluminium oxide supports were used for covalent immobilization of concavalin A, after previous activation with APS (Biotechnology and Applied Biochemistry 16, 221-227 (1992)). Ballardur et al., J. of Colloid and Interface Science, 194, 408-418, 1997 describe the adsorption of oligonucleotides on aminopropyl silane-modified silica wafers. These and similarly loaded metal oxide supports are thus useful for instance in probe-based assays, further for carrying biomolecules, e.g. for use as vaccines, and for separating other substances from mixtures by hybridizing, binding or interacting otherwise with those other substances.

A disadvantage of such supports is that a number of amino-groups of the silanating agent comprising an amine group used for the activation of the metal oxide surface, are still present as unloaded amino-groups even after the support has been loaded with the appropriate biomolecule. This may result in unwanted interactions (non-specific or a-specific interactions) of these amino-groups with various substances present in the medium in which the loaded support is used. For example, when used in probe-based assays, such a-specific interactions may generate high background signals disturbing the signals of the analyte, bound to the capture-biomolecules.

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It has now been found that when the method of the invention is applied, the non-loaded amino-groups are removed selectively from the surface of the further loaded support without affecting the loaded part of the surface.

5 The invention thus relates a method for the preparation of metal oxide supports loaded with biomolecules, comprising the steps of: (a) activating the surface of the support by means of a silanating agent comprising an amine group; (b) loading the support by attaching biomolecules to the activated surface, characterized in that subsequently the loaded support is treated with an acidic solution, and provided that the method is not used for the preparation of silica wafers which are aminated by silanation using (3-aminopropyl)monoethoxydimethylsilane, and loaded
10 with oligonucleotides. Such loaded silica wafers have been subject to a study regarding the effect of pH on the maximum adsorbed amount of oligonucleotides onto the aminated silica wafer [Balladur et al., J. of Colloid and Interface Science, 194, 408-418, 1997]. Therefore this invention does not relate to (a method for the preparation of) such silica wafers.

15 According to another embodiment of the invention the activated and loaded support is subsequently treated with a basic solution (preferably for a prolonged period of time), provided that the method is not used for derivatization of aluminiumoxide nanoparticles aminated with (3-aminopropyl)triethoxysilane, wherein the basic solution further contains a large excess of N-acetylhomocysteinellactone. Such derivatized aluminiumoxide nanoparticles were prepared without the intention to remove aminogroups (Bioconjugate Chem. 1997, 8, 424-433) and are
20 not an embodiment of the present invention (nor is the method for their preparation).

According to another embodiment of the invention the activated and loaded support is subsequently treated with a neutral solution (preferably for a prolonged period of time).

25 The usual methods for suppressing a-specific interactions of unloaded amino-groups on different types of supports involve chemical "capping" / "blocking" the unloaded amino-groups or prehybridizing them with DNA (see for example: Science, Vol. 270, 1995, 467-470; Nucleic Acids Research, Vol. 15 (13) 1987, 5353--5373; Nucleic Acids Research, Vol. 26 (17) 1998, 3883-3891, etc.). However, those methods are not very well defined and require more difficult procedures. Furthermore, with those methods the unloaded amino-groups are not actually
30 removed from the surface, which is the case when the method of the present invention is applied.

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The method of the invention is well defined and easy to perform and the results, e.g. in terms of signal/background ratio's, are comparable or even better when compared to the usual procedures.

5 In any situation in which unloaded amino-groups are to be selectively removed from metal oxide supports loaded with biomolecules, the method of the present invention is applicable, provided that a silanating agent comprising an amine group has been used to activate the metal oxide surface.

10 The pH value of the solution with which the loaded supports may be treated according to the invention, depends substantially on the type of biomolecule attached to the surface and on the way of attaching (loading) the biomolecules to the surface, either covalently or adsorptively.

When the biomolecule is attached to the surface covalently (i.e. they are attached to the amino-groups on the surface via some chemical linkage) and it can withstand both acidic and basic conditions, both acidic and basic solutions may be used for the removal of unloaded amino-
15 groups. If the biomolecule is sensitive to either acidic or basic conditions, respectively a basic or acidic solution must be chosen for the removal of the unloaded amino-groups.

When adsorptively attached (i.e. the biomolecule adheres to the the surface modified with a silanating agent comprising an amine group), also the overall charge of the biomolecule is a
20 relevant parameter. For example, when a negatively charged biomolecule is adsorptively attached to the activated surface of the metal oxide support, it may become detached at a certain pH when the charge of either the biomolecule is reduced by protonation or the charge of the surface is reduced due to deprotonation. For example, an oligonucleotide having a negatively charged backbone will be protonated at very low pH values and it will subsequently
25 be released from the positively charged activated surface of the support. Similarly, the loaded amino-groups are deprotonated at very high pH values (e.g. 11.5) resulting in the release of the negatively charged oligonucleotide.

A person skilled in the art will know how to find the balance between effective removal of the unloaded amino-groups and the intactness of the loaded support. Some preliminary
30 experimentation may be required to find the optimal conditions for the specific circumstances. However, use of an acidic solution is preferred, since the removal of unloaded amino-groups

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The surface of the metal oxide support may be activated using different types of silanating agent comprising an amine group. Effective activating reagents are for example 3-aminopropyl triethoxysilane, 4-aminobutyl-dimethyl-methoxysilane, 3-[2-(2-aminoethylamino)ethylamino]-propyltrimethoxysilane, 3-(2-aminoethylamino)propyl-methyldimethoxysilane, 3-(2-aminoethylamino)propyltrimethoxysilane, 3-aminopropyl-methyl-diethoxysilane, (3-aminopropyl)-tris[2-(2-methoxyethoxy)ethoxy]silane and 4-aminobutyltriethoxysilane, however also other silanating agents comprising an amine group may be equally suitable. A preferred silanating agent is aminopropyl triethoxysilane (APS) used in an unbuffered aqueous solution.

The silanation reaction may be performed in aqueous solution, in organic solution or in the gas phase.

Biomolecules may be attached to the support either covalently or adsorptively. In a preferred embodiment of the invention the biomolecules are adsorptively attached to the activated surface of the support. Preferably, the biomolecules are attached to the surface in spots, thereby forming a (micro)array of spots. A preferred method to spot the surface with biomolecules applies inkjet technology. This technology allows for the accurate deposition of defined volumes of liquid. (See e.g. T.P. Theriault: DNA diagnostic systems based on novel Chem-Jet technologies, IBC Conference on Biochip Array Technologies, Washington DC, May 10, 1995)

In a highly preferred embodiment of the invention an acidic solution of pH 5 is used for the preparation of APS-activated aluminium oxide porous membranes (which are electrochemically manufactured), spotted with arrays of oligonucleotides, which oligonucleotides are adsorptively attached to the activated surface.

Metal oxide supports prepared according to the method of the invention, are very useful for diagnostic purposes, for example in probe-based assays. In particular (micro)arrays, on the surface of which essentially no free amino groups are present that lead to a-specific binding, are very suitable for that purpose. In such arrays, the density of amino groups on the surface area between the spots is significantly lower than the density of amino groups in the spots. Preferably, the arrays comprise different biomolecules in different spots, allowing multi-analyte

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detection. An analyte is a substance, usually a biomolecule, in a mixture which may be detected because of its capability to interact specifically with a selected reagents (for example another biomolecule capable of reacting with the analyte) (on a support).

Probe-based assays comprise for example nucleic acid hybridization assays and immunological assays. In such assays, a sample which comprises an analyte is contacted with a loaded support prepared according to the invention. The analyte is subsequently allowed to bind to the biomolecule which is attached to the surface of the support. Detection of binding can be performed by (1) adding a detection means, for example a substance capable of binding to the analyte, which substance is provided with a label, (2) allowing the detection means to bind to the complex of the analyte and the biomolecule, and (3) determining whether the label is present at the position where the biomolecule was attached. Alternatively, the analyte may already have been provided with a label, in which case binding to the biomolecule can be detected directly, without the addition of a detection means.

The present invention therefore also relates to a kit of parts comprising a metal oxide support according to the invention, further comprising a detection means for determining whether binding has occurred between the biomolecules and an analyte. Preferably, such detection means is a substance capable of binding to the analyte and being provided with a label. Such label is in particular useful, if it is capable of inducing a colour reaction and/or capable of bio- or chemo- or photoluminescence.

Biomolecules which may be used in the process of the invention include oligonucleotides and other negatively charged capture molecules, such as antibodies, antigens, peptides, receptors, haptens and ligands for receptors (which may be modified to introduce (additional) negative charges). Preferred are oligonucleotides. However, the scope of the invention is not limited to these specific molecules. The method may very well be useful to supports loaded with other types of molecules as well.

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The invention is further illustrated by the following examples.

EXAMPLES

Preparation of APS-membranes:

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Membranes (Anodisc 25, 0.2 μ m; Whatman) were silanated by placing them in a 1 % solution of 3-aminopropyltriethoxysilane (Acros, APS) in demineralised water (milliQ), for a period of 1 hr on a plateshaker. The membranes were soaked in water to remove the bulk of the APS molecules and subsequently washed individually by sucking 5 ml of milliQ through the membranes to remove the last traces of free APS. These membranes were heated for 2 hr's at 120 °C under vacuum. After cooling, these "APS-membranes" were stored in a vacuum exsiccator, under argon over KOH until they were used.

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Stability study of APS-membranes:

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The following buffers were made:

- 40 mM Acetate buffer pH 5.0; from sodiumacetate and acetic acid
- 40 mM Acetate buffer pH 6.0; from sodiumacetate and acetic acid
- 40 mM Phosphate buffer pH 7.0; from sodiumdihydrogenphosphate and NaOH
- 20 • 40 mM Borate buffer pH 8.0; from sodium tetraborate and hydrogen chloride
- 40 mM Borate buffer pH 9.0; from sodium tetraborate and hydrogen chloride

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The APS membranes were incubated at the above mentioned pH's using the appropriate buffer for a period between 0.25 and 22 hr's (see table 1, for details). After this period the membrane was removed from the buffer solution and washed with milliQ and ethanol and dried under vacuum at room temperature. Subsequently the amount of amino-groups was determined using a modified trinitrobenzenesulfonic acid (TNBS) method [Habeeb, A.F.S.A. (1996), Anal. Biochem. 14, 328. See also Sashidhar, R.B., Capoor, A.K., Ramana, D., (1994), J. Immunol. M]. A commercially available stock solution (Sigma) of 5% (m/v) of TNBS was diluted 25-fold with a 50 mM borate buffer of pH 10.0 directly before the amino-group determination was performed. Each treated membrane was placed individually in a beaker and 4 ml of the diluted

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TNBS solution was added and the membrane was incubated on a plateshaker for 1 hr at room temperature. The yellow/orange membranes were thoroughly washed (by using vacuum to suck liquids through the membrane) with milliQ to remove excess TNBS. Subsequently the membranes were dissolved in a known amount (e.g. 4.0 ml) of a 4.0 M NaOH solution in milliQ. After filtration of this solution over a 0.45 μ m filter, the absorption was measured using an UV/vis spectrophotometer. Using the extinction coefficient of the adduct at 390 nm of 11929 l/mol/cm, the number of amino groups per membrane that reacted with TNBS could be determined. In table 1 the decrease (in %) in amino-groups as function of time and pH is given.

Time (hr)	Percentage of aminogroups at				
	pH 5.0	pH 6.0	pH 7.0	pH 8.0	pH 9.0
Blanc exp.	100	100	100	100	100
0.25	54				
0.5	19				
1	9				
1.5		49			
2.5		17	82	96	51
6			69	52	34
22			54	42	21

Table 1: degradation as function of pH and time of APS membranes

From this table it is clear that the loss of aminogroups is fastest at lower pH's.

Protective effects of loading of the activated surface of a support:

(a) Protective effect of the TNBS group:

In another experiment the aminogroups on an APS membrane reacted with TNBS as described above. This yielded membranes that contain covalently linked chromophores that consist of the trinitrobenzene adduct of the amino group (APS-TNB membranes).

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These membranes were treated with the same buffers mentioned above for a period of 22 hr's at room temperature. In table 2 the remaining amount of chromophores after this period, as determined with the above mentioned method, are summarized.

pH applied	Resulting aminogroups eq chromophores (%)
Blanc experiment	100
5.0	71
6.0	95
7.0	83
8.0	90
9.0	95

5 Table 2: stability of TNB-APS membranes

This table clearly shows, that functionalisation of the aminogroups results in a large increase of the stability. In other words, the majority of the functionalized aminogroups attached to the surface is not affected.

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(b) Protective effect of DNA

A haring sperm DNA solution of 1 µg/ml in milliQ was prepared and 1 µl of this solution was pipetted onto the APS membranes in spots. These membranes were divided in two sets: one which was treated with pH 5 buffer (for 1 hr) and the other which were not. Both sets of membranes were subsequently treated with TNBS (method described above) in order to detect any decrease in amino-groups. This yielded the following results. The total surface of the untreated membrane showed the expected orange colour since no degradation (removal) of APS groups had taken place. The surface of the membrane that was treated at pH 5.0 was not orange but very slightly yellow, due to the degradation of nearly all APS molecules. However, at the spots where the DNA was pipetted on the membrane, an orange colour was still visible, indicating the protective effect of the DNA on the degradation process of the aminogroups on the surface. It was shown in a blanc experiment that the amino groups of DNA did not react

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with TNBS in such a way that this could explain the orange colour observed for the pH 5 treated membrane.

(c) Protective effect of oligonucleotides

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A solution of 200uM oligonucleotide with the sequence 5'ATAT GGC TCT CCC GGG AGGGGG GGT CCT GGA 3' was prepared. This solution was pipetted onto two unmodified membranes and onto two APS membranes. In this process only one half of every membrane was covered with the oligonucleotide. Subsequently these membranes were dried at 37 °C for one hr. One unmodified and one APS membrane were treated with pH 5 solution as described above. Subsequently all membranes were washed with phosphate buffer pH 7.4 and ethanol and dried. All membranes were placed in the diluted TNBS solution as described above and incubated for one hr. After washing and drying, the unmodified membranes were totally white, while the total surface of the APS membrane that was not treated with pH 5 was yellow.

15 The pH 5 treated APS membrane showed a slightly yellow color on the half that was covered with the oligonucleotides while the other half was totally white.

This experiments clearly shows the protective effect of oligonucleotides.

(d) Arrays of oligonucleotides:

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The protective effect was also visualised in a different way using hybridization of oligonucleotides:

Using a commercially available spotter (Packard instruments), droplets of 300 picoliter of oligonucleotide A (100 μ M dissolved in milliQ) were spotted onto APS membranes:

25 This oligonucleotide **A** is defined by the sequence: 5'-TTG TAC AGA ACT GGA AAA GGA
3'. Also available for hybridization experiments is oligonucleotide **B** (sequence 5'-TCC TTT
TCC AGT TCT GTA CAA 3') which is complementary to **A** and labeled with a fluoresceine
moiety at the 5' end.

30 After spotting, all membranes were dried at 50 °C for 1 hr. One set of membranes was incubated in an acetate buffer (40 mM, pH 5.0) for 30 minutes and subsequently washed with

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milliQ, ethanol, and dried under vacuum. These membranes will be referred to as "pH 5 arrays". As a blanc experiment the other set of membranes was not treated with the pH 5.0 solution, but it was only washed with milliQ, ethanol and dried under vacuum. These membranes will be called "blanc array". All membranes were stored under argon until being used for hybridisation experiments.

The membranes were placed in a homebuilt device that allows 25 μ l of a phosphate buffer (pH 7.0) containing the complementary oligonucleotide B being flushed back and forth through the membrane. If the solution is below this device, a hybridised nucleotide on the membrane can directly be seen due to its fluorescent label using an ordinary fluorescence microscope and a CCD-camera. With this setup, the data in table 3 have been collected. One cycle is defined as the 25 μ l being pumped up and down one time through the membrane. Specific signals have been corrected for the background signal.

Signal (arb.units) for pH 5 arrays			Signal (arb.units) for blanc arrays		
Cycles	Specific	Background	Cycles	Specific	Background
0	0	5	0	0	5
2	4	8	1	0	41
5	7	8	5	2	47
10	11	9			
15	13	9			
20	15	9			

Table 3: Specific and a-specific interactions of pH 5 arrays and blanc arrays

From this table the following observations are important:

- The background signal for pH 5 arrays is low and nearly constant
- The specific hybridisation for pH 5 arrays takes place rapidly and can easily be monitored

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- The blanc array does not show specific signals, due to a strong α -specific interaction between the amino-groups (which are present all over the membrane) and oligonucleotide B.
- The α -specific interactions (background) that are seen for the blanc arrays, is highly reduced by the acidic treatment.

Thus, this experiment shows that the acidic treatment removes all amino groups that are not covered and protected by oligonucleotides. Oligonucleotide A is not removed from the surface indicating that it is still captured by aminogroups on the surface below the nucleotides which have not been removed.